

Optimization of cholinesterase-based catalytic bioscavengers against organophosphorus agents

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Abstract

© 2018 Lushchekina, Schopfer, Grigorenko, Nemukhin, Varfolomeev, Lockridge and Masson. Organophosphorus agents (OPs) are irreversible inhibitors of acetylcholinesterase (AChE). OP poisoning causes major cholinergic syndrome. Current medical counter-measures mitigate the acute effects but have limited action against OP-induced brain damage. Bioscavengers are appealing alternative therapeutic approach because they neutralize OPs in bloodstream before they reach physiological targets. First generation bioscavengers are stoichiometric bioscavengers. However, stoichiometric neutralization requires administration of huge doses of enzyme. Second generation bioscavengers are catalytic bioscavengers capable of detoxifying OPs with a turnover. High bimolecular rate constants ($k_{cat}/K_m > 10^6 \text{ M}^{-1} \text{ min}^{-1}$) are required, so that low enzyme doses can be administered. Cholinesterases (ChE) are attractive candidates because OPs are hemi-substrates. Moderate OP hydrolase (OPase) activity has been observed for certain natural ChEs and for G117H-based human BChE mutants made by site-directed mutagenesis. However, before mutated ChEs can become operational catalytic bioscavengers their dephosphorylation rate constant must be increased by several orders of magnitude. New strategies for converting ChEs into fast OPase are based either on combinational approaches or on computer redesign of enzyme. The keystone for rational conversion of ChEs into OPases is to understand the reaction mechanisms with OPs. In the present work we propose that efficient OP hydrolysis can be achieved by re-designing the configuration of enzyme active center residues and by creating specific routes for attack of water molecules and proton transfer. Four directions for nucleophilic attack of water on phosphorus atom were defined. Changes must lead to a novel enzyme, wherein OP hydrolysis wins over competing aging reactions. Kinetic, crystallographic, and computational data have been accumulated that describe mechanisms of reactions involving ChEs. From these studies, it appears that introducing new groups that create a stable H-bonded network susceptible to activate and orient water molecule, stabilize transition states (TS), and intermediates may determine whether dephosphorylation is favored over aging. Mutations on key residues (L286, F329, F398) were considered. QM/MM calculations suggest that mutation L286H combined to other mutations favors water attack from apical position. However, the aging reaction is competing. Axial direction of water attack is not favorable to aging. QM/MM calculation shows that F329H+F398H-based multiple mutants display favorable energy barrier for fast reactivation without aging.

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Keywords

Acetylcholinesterase, Bioscavenger, Butyrylcholinesterase, Computer design,

References

- [1] Albaret, C., Masson, P., Broomfield, C. A., El Kaim, L., and Fortier, P. L. (1998). 'Mechanical aspects of the phosphotriesterase activity of human butyrylcholinesterase G117H mutant'. in *Structure and Function of Cholinesterases and Related Proteins*, 1st Edn., eds B. P. Doctor, D. M. Quinn, P. Taylot, and R. L. Rotundo (New York, NY: Plenum Press), 399-405
- [2] Amitay, M., and Shurki, A. (2009). The structure of G117H mutant of butyrylcholinesterase: nerve agents scavenger. *Proteins* 77, 370-377. doi: 10.1002/prot.22442
- [3] Amitay, M., and Shurki, A. (2011). Hydrolysis of organophosphate compounds by mutant butyrylcholinesterase: a story of two histidines. *Proteins* 79, 352-364. doi: 10.1002/prot.22864
- [4] Ashani, Y., Leader, H., Aggarwal, N., Silman, I., Worek, F., Sussman, J. L., et al. (2016). In vitro evaluation of the catalytic activity of paraoxonases and phosphotriesterases predicts the enzyme circulatory levels required for in vivo protection against organophosphate intoxications. *Chem. Biol. Interact.* 259(Pt B), 252-256. doi: 10.1016/j.cbi.2016.04.039
- [5] Bigley, A. N., Mabanglo, M. F., Harvey, S. P., and Raushel, F. M. (2015). Variants of phosphotriesterase for the enhanced detoxification of the chemical warfare agent VR. *Biochemistry* 54, 5502-5512. doi: 10.1021/acs.biochem.5b00629
- [6] Brazzolotto, X., Froment, M. T., Gessay, F., Worek, F., Dorandeu, F., and Nachon, F. (2015). 'Biochemical and structural study of a self-reactivating butyrylcholinesterase after V-type nerve agent inhibition'. in *12th International Meeting of Cholinesterases-6th-Paraoxonase Conference* (Elche)
- [7] Brazzolotto, X., Igert, A., Guillon, V., Santoni, G., and Nachon, F. (2017). Bacterial expression of human butyrylcholinesterase as a tool for nerve agent bioscavengers development. *Molecules* 22:1828. doi: 10.3390/molecules22111828
- [8] Broomfield, C. A., Lockridge, O., and Millard, C. B. (1999). Protein engineering of a human enzyme that hydrolyzes V and G nerve agents: design, construction and characterization. *Chem. Biol. Inter.* 119-120, 413-418. doi: 10.1016/s0009-2797(99)00053-8
- [9] Dafferner, A. J., Lushchekina, S., Masson, P., Xiao, G., Schopfer, L. M., and Lockridge, O. (2017). Characterization of butyrylcholinesterase in bovine serum. *Chem. Biol. Interact.* 266, 17-27. doi: 10.1016/j.cbi.2017.02.004
- [10] Doctor, B. P., and Saxena, A. (2005). Bioscavengers for the protection of humans against organophosphate toxicity. *Chem. Biol. Interact.* 157-158, 167-171. doi: 10.1016/j.cbi.2005.10.024
- [11] Dorandeu, F., Foquin, A., Briot, R., Delacour, C., Denis, J., Alonso, A., et al. (2008). An unexpected plasma cholinesterase activity rebound after challenge with a high dose of the nerve agent VX. *Toxicology* 248, 151-157. doi: 10.1016/j.tox.2008.03.013
- [12] Eddleston, M., Buckley, N. A., Eyer, P., and Dawson, A. H. (2008). Management of acute organophosphorus pesticide poisoning. *Lancet* 371, 597-607. doi: 10.1016/S0140-6736(07)61202-1
- [13] Field, M. J., and Wymore, T. W. (2014). Multiscale modeling of nerve agent hydrolysis mechanisms: a tale of two nobel prizes. *Phys. Scrip.* 89:108004. doi: 10.1088/0031-8949/89/10/108004
- [14] Goldenzweig, A., Goldsmith, M., Hill, S. E., Gertman, O., Laurino, P., Ashani, Y., et al. (2016). Automated structure-and sequence-based design of proteins for high bacterial expression and stability. *Mol. Cell* 63, 337-346. doi: 10.1016/j.molcel.2016.06.012
- [15] Goldsmith, M., Aggarwal, N., Ashani, Y., Jubran, H., Greisen, P. J., Ovchinnikov, S., et al. (2017). Overcoming an optimization plateau in the directed evolution of highly efficient nerve agent bioscavengers. *Protein Eng. Des. Sel.* 30, 333-345. doi: 10.1093/protein/gzx003
- [16] Goldsmith, M., Eckstein, S., Ashani, Y., Greisen, P. Jr., Leader, H., Sussman, J. L., et al. (2016). Catalytic efficiencies of directly evolved phosphotriesterase variants with structurally different organophosphorus compounds in vitro. *Arch. Toxicol.* 90, 2711-2724. doi: 10.1007/s00204-015-1626-2
- [17] Hiblot, J., Bzdrenga, J., Champion, C., Chabriere, E., and Elias, M. (2015). Crystal structure of VmoLac, a tentative quorum quenching lactonase from the extremophilic crenarchaeon *Vulcanisaeta moutnovskia*. *Sci. Rep.* 5:8372. doi: 10.1038/srep08372
- [18] Jackson, C. J., Liu, J.-W., Carr, P. D., Younus, F., Coppin, C., Meirelles, T., et al. (2013). Structure and function of an insect α -carboxylesterase (α Esterase7) associated with insecticide resistance. *Proc. Natl. Acad. Sci. U.S.A.* 110, 10177-10182. doi: 10.1073/pnas.1304097110
- [19] Jacob, R. B., Michaels, K. C., Anderson, C. J., Fay, J. M., and Dokholyan, N. V. (2016). Harnessing nature's diversity: discovering organophosphate bioscavenger characteristics among low molecular weight proteins. *Sci. Rep.* 6:37175. doi: 10.1038/srep37175

- [20] Jacquet, P., Daudé, D., Bzdrenga, J., Masson, P., Elias, M., and Chabrière, E. (2016). Current and emerging strategies for organophosphate decontamination: special focus on hyperstable enzymes. *Environ. Sci. Pollut. Res.* 23, 8200-8218. doi: 10.1007/s11356-016-6143-1
- [21] Jarv, J. (1984). Stereochemical aspects of cholinesterase catalysis. *Bioorg. Chem.* 12, 259-278. doi: 10.1016/0045-2068(84)90010-5
- [22] Kovarik, Z., Macek Hrvat, N., Katalinic, M., Sit, R. K., Paradyse, A., Zunec, S., et al. (2015). Catalytic soman scavenging by the Y337A/F338A acetylcholinesterase mutant assisted with novel site-directed aldoximes. *Chem. Res. Toxicol.* 28, 1036-1044. doi: 10.1021/acs.chemrestox.5b00060
- [23] Kulakova, A., Lushchekina, S., Grigorenko, B., and Nemukhin, A. (2015). Modeling reactivation of the phosphorylated human butyrylcholinesterase by QM(DFTB)/MM calculations. *J. Theor. Comp. Chem.* 14:1550051. doi: 10.1142/s0219633615500510
- [24] Kumler, W. D., and Eiler, J. J. (1943). The acid strength of mono and diesters of phosphoric acid. The n-alkyl esters from methyl to butyl, the esters of biological importance, and the natural guanidine phosphoric acids. *J. Am. Chem. Soc.* 65, 2355-2361. doi: 10.1021/ja01252a028
- [25] Lenz, D. E., Maxwell, D. M., Koplovitz, I., Clark, C. R., Capacio, B. R., Cerasoli, D. M., et al. (2005). Protection against soman or VX poisoning by human butyrylcholinesterase in guinea pigs and cynomolgus monkeys. *Chem. Biol. Interact.* 157-158, 205-210. doi: 10.1016/j.cbi.2005.10.025
- [26] Lockridge, O. (2015). Review of human butyrylcholinesterase structure, function, genetic variants, history of use in the clinic, and potential therapeutic uses. *Pharmacol. Ther.* 148, 34-46. doi: 10.1016/j.pharmthera.2014.11.011
- [27] Lockridge, O., Blong, R. M., Masson, P., Froment, M. T., Millard, C. B., and Broomfield, C. A. (1997). A single amino acid substitution, Gly117His, confers phosphotriesterase (organophosphorus acid anhydride hydrolase) activity on human butyrylcholinesterase. *Biochemistry* 36, 786-795. doi: 10.1021/bi961412g
- [28] Mabbitt, P. D., Correy, G. J., Meirelles, T., Fraser, N. J., Coote, M. L., and Jackson, C. J. (2016). Conformational disorganization within the active site of a recently evolved organophosphate hydrolase limits its catalytic efficiency. *Biochemistry* 55, 1408-1417. doi: 10.1021/acs.biochem.5b01322
- [29] Massiah, M. A., Viragh, C., Reddy, P. M., Kovach, I. M., Johnson, J., Rosenberry, T. L., et al. (2001). Short, strong hydrogen bonds at the active site of human acetylcholinesterase: proton NMR studies. *Biochemistry* 40, 5682-5690. doi: 10.1021/bi010243j
- [30] Masson, P. (2016a). 'Nerve agents: catalytic scavengers, alternative approach for medical countermeasures'. in *Chemical Warfare Toxicology*, eds F. Worek, J. Jenner, and H. Thiermann (Cambridge, UK: Royal Society of Chemistry Pub), 43-81
- [31] Masson, P. (2016b). Novel approaches in prophylaxis/pretreatment and treatment of organophosphorus poisoning. *Phosph. Sulfur Silicon Relat. Elements* 191, 1433-1443. doi: 10.1080/10426507.2016.1211652
- [32] Masson, P., Fortier, P.-L., Albaret, C., Froment, M.-T., Bartels, C. F., and Lockridge, O. (1997a). Aging of diisopropyl-phosphorylated human butyrylcholinesterase. *Biochem. J.* 327, 601-607. doi: 10.1042/bj3270601
- [33] Masson, P., Froment, M. T., Bartels, C. F., and Lockridge, O. (1997b). Importance of aspartate-70 in organophosphate inhibition, oxime re-activation and aging of human butyrylcholinesterase. *Biochem. J.* 325 (Pt 1), 53-61
- [34] Masson, P., Legrand, P., Bartels, C. F., Froment, M.-T., Schopfer, L. M., and Lockridge, O. (1997c). Role of aspartate 70 and tryptophan 82 in binding of succinylthiocholine to human butyrylcholinesterase. *Biochemistry* 36, 2266-2277. doi: 10.1021/bi962484a
- [35] Masson, P., and Lushchekina, S. V. (2016). Emergence of catalytic bioscavengers against organophosphorus agents. *Chem. Biol. Interact.* 259(Pt B), 319-326. doi: 10.1016/j.cbi.2016.02.010
- [36] Masson, P., and Nachon, F. (2017). Cholinesterase reactivators and bioscavengers for pre-and post-exposure treatments of organophosphorus poisoning. *J. Neurochem.* 142(Suppl. 2), 26-40. doi: 10.1111/jnc.14026
- [37] Masson, P., Nachon, F., Broomfield, C. A., Lenz, D. E., Verdier, L., Schopfer, L. M., et al. (2008). A collaborative endeavor to design cholinesterase-based catalytic scavengers against toxic organophosphorus esters. *Chem. Biol. Interact.* 175, 273-280. doi: 10.1016/j.cbi.2008.04.005
- [38] Masson, P., Nachon, F., and Lockridge, O. (2010). Structural approach to the aging of phosphorylated cholinesterases. *Chem. Biol. Interact.* 187, 157-162. doi: 10.1016/j.cbi.2010.03.027
- [39] Millard, C. B., Lockridge, O., and Broomfield, C. A. (1995). Design and expression of organophosphorus acid anhydride hydrolase activity in human butyrylcholinesterase. *Biochemistry* 34, 15925-15933. doi: 10.1021/bi00049a007
- [40] Millard, C. B., Lockridge, O., and Broomfield, C. A. (1998). Organophosphorus acid anhydride hydrolase activity in human butyrylcholinesterase: synergy results in a somanase. *Biochemistry* 37, 237-247. doi: 10.1021/bi972057c
- [41] Mumford, H., Docx, C. J., Price, M. E., Green, A. C., Tattersall, J. E. H., and Armstrong, S. J. (2013). Human plasma-derived BuChE as a stoichiometric bioscavenger for treatment of nerve agent poisoning. *Chem. Biol. Interact.* 203, 160-166. doi: 10.1016/j.cbi.2012.08.018

- [42] Myhrer, T., and Aas, P. (2016). Pretreatment and prophylaxis against nerve agent poisoning: are undesirable behavioral side effects unavoidable? *Neurosci. Biobehav. Rev.* 71, 657-670. doi: 10.1016/j.neubiorev.2016.10.017
- [43] Nachon, F., Asojo, O. A., Borgstahl, G. E., Masson, P., and Lockridge, O. (2005). Role of water in aging of human butyrylcholinesterase inhibited by echothiophate: the crystal structure suggests two alternative mechanisms of aging. *Biochemistry* 44, 1154-1162. doi: 10.1021/bi048238d
- [44] Nachon, F., Brazzolotto, X., Trovaslet, M., and Masson, P. (2013). Progress in the development of enzyme-based nerve agent bioscavengers. *Chem. Biol. Interact.* 206, 536-544. doi: 10.1016/j.cbi.2013.06.012
- [45] Nachon, F., Carletti, E., Wandhammer, M., Nicolet, Y., Schopfer, L. M., Masson, P., et al. (2011). X-ray crystallographic snapshots of reaction intermediates in the G117H mutant of human butyrylcholinesterase, a nerve agent target engineered into a catalytic bioscavenger. *Biochem. J.* 434, 73-82. doi: 10.1042/bj20101648
- [46] Nemukhin, A. V., Kulakova, A. M., Lushchekina, S. V., Ermilov, A. Y., and Varfolomeev, S. D. (2015). Modeling chemical transformations at the active sites of cholinesterases by quantum-based simulations. *Mosc. Univ. Chem. Bull.* 70, 274-277. doi: 10.3103/S0027131415060061
- [47] Nicolet, Y., Lockridge, O., Masson, P., Fontecilla-Camps, J. C., and Nachon, F. (2003). Crystal structure of human butyrylcholinesterase and of its complexes with substrate and products. *J. Biol. Chem.* 278, 41141-41147. doi: 10.1074/jbc.M210241200
- [48] Onder, S., David, E., Tacal, O., Schopfer, L. M., and Lockridge, O. (2017). Hupresin retains binding capacity for butyrylcholinesterase and acetylcholinesterase after sanitation with sodium hydroxide. *Front. Pharmacol.* 8:713. doi: 10.3389/fphar.2017.00713
- [49] Pashirova, T. N., Zueva, I. V., Petrov, K. A., Babaev, V. M., Lukashenko, S. S., Rizvanov, I. K., et al. (2017). Nanoparticle-delivered 2-PAM for rat brain protection against paraoxon central toxicity. *ACS Appl. Mater. Interfaces* 9, 16922-16932. doi: 10.1021/acsami.7b04163
- [50] Patocka, J. (2017). What killed Kim Jong-nam? Was it the agent VX. *Mil. Med. Sci. Lett.* 86, 1-4
- [51] Peng, H., Brimijoin, S., Hrabovska, A., Targosova, K., Krejci, E., Blake, T. A., et al. (2015). Comparison of 5 monoclonal antibodies for immunopurification of human butyrylcholinesterase on Dynabeads: KD values, binding pairs, and amino acid sequences. *Chem. Biol. Interact.* 240, 336-345. doi: 10.1016/j.cbi.2015.08.024
- [52] Poyot, T., Nachon, F., Froment, M. T., Loiodice, M., Wieseler, S., Schopfer, L. M., et al. (2006). Mutant of *Bungarus fasciatus* acetylcholinesterase with low affinity and low hydrolase activity toward organophosphorus esters. *Biochim. Biophys. Acta* 1764, 1470-1478. doi: 10.1016/j.bbapap.2006.07.008
- [53] Radic, Z., Dale, T., Kovarik, Z., Berend, S., Garcia, E., Zhang, L., et al. (2013). Catalytic detoxification of nerve agent and pesticide organophosphates by butyrylcholinesterase assisted with non-pyridinium oximes. *Biochem. J.* 450, 231-242. doi: 10.1042/bj20121612
- [54] Restaino, O. F., Borzacchiello, M. G., Scognamiglio, I., Porzio, E., Manco, G., Fedele, L., et al. (2017). Boosted large-scale production and purification of a thermostable archaeal phosphotriesterase-like lactonase for organophosphate decontamination. *J. Ind. Microbiol. Biotechnol.* 44, 363-375. doi: 10.1007/s10295-016-1892-x
- [55] Rice, H., Mann, T. M., Armstrong, S. J., Price, M. E., Green, A. C., and Tattersall, J. E. (2016). The potential role of bioscavenger in the medical management of nerve-agent poisoned casualties. *Chem. Biol. Interact.* 259(Pt B), 175-181. doi: 10.1016/j.cbi.2016.04.038
- [56] Schopfer, L. M., Boeck, A. T., Broomfield, C. A., and Lockridge, O. (2004). Mutants of human butyrylcholinesterase with organophosphate hydrolase activity; evidence that His117 is a general base catalyst for hydrolysis of echothiophate. *J. Med. Chem. Biol. Radiol. Def.* 2, 1-21
- [57] Steitz, T. A., and Shulman, R. G. (1982). Crystallographic and NMR studies of the serine proteases. *Annu. Rev. Biophys. Bioeng.* 11, 419-444. doi: 10.1146/annurev.bb.11.060182.002223
- [58] Terekhov, S. S., Smirnov, I. V., Stepanova, A. V., Bobik, T. V., Mokrushina, Y. A., Ponomarenko, N. A., et al. (2017). Microfluidic droplet platform for ultrahigh-throughput single-cell screening of biodiversity. *Proc. Natl. Acad. Sci. U.S.A.* 114, 2550-2555. doi: 10.1073/pnas.1621226114
- [59] Valiev, M., Bylaska, E. J., Govind, N., Kowalski, K., Straatsma, T. P., Van Dam, H. J. J., et al. (2010). NWChem: a comprehensive and scalable open-source solution for large scale molecular simulations. *Comput. Phys. Commun.* 181, 1477-1489. doi: 10.1016/j.cpc.2010.04.018
- [60] Viragh, C., Harris, T. K., Reddy, P. M., Massiah, M. A., Mildvan, A. S., and Kovach, I. M. (2000). NMR evidence for a short, strong hydrogen bond at the active site of a cholinesterase. *Biochemistry* 39, 16200-16205. doi: 10.1021/bi0022644
- [61] Voevodin, V. V., Zhumatiy, S. A., Sobolev, S. I., Antonov, A. S., Bryzgalov, P. A., Nikitenko, D. A., et al. (2012). Practice of 'Lomonosov' supercomputer. *Open Syst. J.* 7, 36-39
- [62] Wandhammer, M., Carletti, E., van der Schans, M., Gillon, E., Nicolet, Y., Masson, P., et al. (2011). Structural study of the complex stereoselectivity of human butyrylcholinesterase for the neurotoxic V-agents. *J. Biol. Chem.* 286, 16783-16789. doi: 10.1074/jbc.M110.209569

- [63] Wille, T., Neumaier, K., Koller, M., Ehinger, C., Aggarwal, N., Ashani, Y., et al. (2016). Single treatment of VX poisoned guinea pigs with the phosphotriesterase mutant C23AL: intraosseous versus intravenous injection. *Toxicol. Lett.* 258, 198-206. doi: 10.1016/j.toxlet.2016.07.004
- [64] Worek, F., Thiermann, H., and Wille, T. (2016a). Catalytic bioscavengers in nerve agent poisoning: a promising approach? *Toxicol. Lett.* 244, 143-148. doi: 10.1016/j.toxlet.2015.07.012
- [65] Worek, F., Wille, T., Koller, M., and Thiermann, H. (2016b). Toxicology of organophosphorus compounds in view of an increasing terrorist threat. *Arch. Toxicol.* 90, 2131-2145. doi: 10.1007/s00204-016-1772-1
- [66] Yao, Y., Liu, J., and Zhan, C.-G. (2012). Why does the G117H mutation considerably improve the activity of human butyrylcholinesterase against sarin? Insights from quantum mechanical/molecular mechanical free energy calculations. *Biochemistry* 51, 8980-8992. doi: 10.1021/bi3009246